ORIGINAL ARTICLE

Mitochondrial diversity in Amerindian Kichwa and Mestizo populations from Ecuador

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Abstract This study presents mitochondrial DNA (mtDNA) data from 107 unrelated individuals from two of the major ethnic groups in Ecuador: Amerindian Kichwas (n=65) and Mestizos (n=42). We characterized the diversity of the matrilineal lineages of these Ecuadorian groups by analyzing the entire mtDNA control region. Different patterns of diversity were observed in the two groups as result of the unique historical and demographic events which have occurred in each population. Higher genetic diversity values were obtained for the Mestizo group than for the Amerindian group. Interestingly, only Native American lineages were detected in the two population samples, but with differences in the haplogroup distribution: Kichwa (A, 49%; B, 3%; C, 8%; and D, 40%) and Mestizo (A, 33%; B, 33%; C, 10%; and D, 24%). Analysis of the complete mtDNA control region proved to be useful to increase the discrimination power between individuals who

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L. Roewer Department of Forensic Genetics, Institute of Legal Medicine and Forensic Sciences, Charité-Universitätsmedizin Berlin, Berlin, Germany showed common haplotypes in HVSI and HVSII segments; and added valuable information to the phylogenetic interpretation of mtDNA haplotypes.

 $\textbf{Keywords} \ \, \text{mtDNA population data} \cdot \text{Ecuador} \cdot \text{Native} \\ \text{American} \cdot \text{Control region}$

Introduction

Mitochondrial DNA (mtDNA) analysis is a very useful tool for population studies and forensic applications because of its maternal inheritance, rapid rate of evolution, absence of recombination, and high copy number per cell. Most studies have focused on the noncoding region of mtDNA, known as the control region or D-Loop. This region is highly polymorphic and can be used to study short-term evolutionary events and for human forensic identification especially in cases where DNA is degraded or in low quantities [1, 2]. However, the standard typing of the control region, based on the hypervariable segments HVSI and HVSII, can result in a rather limited power of discrimination. In order to overcome this drawback, the analysis of the entire control region and/or the typing of single nucleotide polymorphisms in the coding region seems to be effective approaches [3, 4].

MtDNA studies have been used to reconstruct the human demographic history as well as to infer the ancestry of individuals [5]. In this sense, the study of American human groups has awakened a growing interest because of their complexity and diversity [6]. This is the case of the Ecuadorian population, a multicultural and pluri-ethnic population with three main groups: (a) Mestizos, an admixed population of Spanish and Amerindian descendants (72%); (b) Amerindian natives with more than 13 major groups



(7%), being the Kichwa (Quechua speaking) the most numerous group; and (c) African-derived populations (7%) [7].

Previous studies, based on autosomal and Y chromosome STRs [8, 9], have evidenced variable genetic contribution of European, Amerindian, and African ancestry in the Ecuadorian groups, as a result of the European colonization and the arrival of African slaves during colonial times. Moreover, a certain level of sex-biased genetic admixture has been detected in the Ecuadorian Mestizo population, which has been suggested to be the reflection of the preferential mating that occurred between European male settlers and Native American females. In order to provide the female side of the history, some mtDNA population studies have been carried out, but they have focused on small groups as Waorani [10, 11] and Cayapa [12].

In this work, the study of the mtDNA control region from the two main Ecuadorian ethnic groups has been carried out with the following aims: (a) to study the genetic structure of these groups through mtDNA; (b) to infer the demographic and historical processes that may have influenced the genetic diversity of these groups; and (c) to contribute to mtDNA databases of Ecuadorian populations for forensic purposes.

Materials and methods

The population sample included 107 maternally unrelated individuals born and living in Ecuador: 65 self-identified Amerindian Kichwas from the Amazonian provinces of Pastaza, Orellana, and Napo (Kichwa del Oriente) and 42 Mestizos from different regions of the country. All donors gave their informed consent prior to inclusion in the study.

DNA was extracted from blood stains on FTA cards (Whatman Inc., Clifton, NJ) using the Chelex extraction procedure [13]. The entire control region was amplified from positions 16024 to 576 using the primer sets L15988 and H616 as described in a previous study [14]. DNA products were then purified with ExoSAP (USB) and sequenced using the same set of primers L15988/H616 as well as L16363 [15], L29 [16], and H7 [17]. Sequencing extension reactions were performed using BigDye Termination v3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, CA) following the manufacturer's conditions. Purification of products was completed with IBIAN ® Dye Clean-up (IBIAN Technologies, Spain). Automated DNA sequencing was carried out on the ABI PrismTM 310 Genetic Analyzer (Applied Biosystems).

The sequences were aligned and compared to the revised Cambridge Reference Sequence [18] using the SeqScape software (Applied Biosystems). Statistical analyses were performed using Arlequin v3.11 software [19], ignoring C-stretch variation. Diversity estimations were calculated for the entire mtDNA control region (16024 to 576), although some calculations were carried out for HVSI (16024-16365), HVSII (73-340), and HVSIII (438-574). Exact tests

of population differentiation and pairwise Fst were also performed. Haplogroups were defined on the basis of entire control region sequences, according to the phylogenetic tree of global human mtDNA v12 (available at http://www.phylotree.org/).

Sequences will be searchable via the EMPOP database under accession numbers EMP00421 for the Kichwa and EMP00422 for the Mestizo individuals.

Results and discussion

Diversity in the two Ecuadorian populations

The haplotypes obtained for Kichwa and Mestizo individuals are provided in Table S1. The analysis of the complete mitochondrial control region resulted in 39 distinct haplotypes in the entire Ecuadorian sample, with four sequences common to both ethnic groups. Within the Kichwa subset, 13 different haplotypes were described, with five of them being unique to single individuals. The most frequent sequence was shared by 40% of the Kichwa individuals. In the Mestizo subset, 30 different haplotypes were observed, and 23 of these sequences were present in just one individual. The most frequent sequence in the Mestizo sample set was observed in 14.29% of the cases.

It is noteworthy that when the analysis was restricted to the HVSI and HVSII regions, a lower number of haplotypes could be differentiated when compared to the entire control region analysis, in Kichwa (11 vs 13) and Mestizo (29 vs 30). The analysis of the HVSIII did not allow further differentiation among individuals who shared identical HVSI–HVSII haplotypes. Thus, it was the analysis of non-HVS regions that increased the total number of haplotypes detected.

Other parameters characterizing within-population diversity based on the complete mtDNA control region sequences are listed in Table 1.

Haplotype diversities obtained for the entire control region were 0.8029 ± 0.0373 and 0.9733 ± 0.0144 for the Kichwa and Mestizo groups, respectively. These differences are explained by the higher number of different sequences

Table 1 Diversity measures for control region data from Kichwa and Mestizo populations

	Kichwa	Mestizo
Number of individuals	65	42
Different haplotypes	13	30
Number of polymorphic sites	47	83
Haplotype diversity	$0.8029\!\pm\!0.0373$	$0.9733\!\pm\!0.0144$
Random match probability (%)	0.2095	0.0499
Power of discrimination	0.7905	0.9501



and proportion of single haplotypes over total sequences found in Mestizos.

These results are consistent with general patterns described in other Mestizo and Amerindian populations [3, 12, 20, 21]. Mestizo groups usually exhibit high diversities because they are composed of lineages from different geographical and ethnic origins, which results in a high heterogeneity in their genetic pool. On the other hand, Amerindian populations normally present a lower diversity range, with values that are especially lower in groups that have suffered restricted gene flow and high inbreeding levels over long periods [10, 22]. The intermediate diversity values described for the Kichwa group seem to indicate that this group did not experience important bottlenecks or genetic drifts and maintained its heterogeneity through gene flow from other groups.

Furthermore, to evaluate the usefulness of mtDNA analysis for forensic purposes in these populations, the probability of randomly selected sequences matching (RMP) was estimated, which is inversely proportional to the power of discrimination of a genetic system. In a logical reflection of the data on diversity parameters, the RMP value was smaller in Mestizo population (5%) than in Kichwa (20.95%). Accordingly to these results, the power of discrimination of the mtDNA in forensic identity testing would be more limited for the Amerindian population, where it would more probable to find two individuals with the same haplotype.

Comparison of the two Ecuadorian populations

The exact test of population differentiation based on haplotype frequencies showed significant differences for mtDNA sequences between the two Ecuadorian populations (P<0.001). The same results were obtained through Fst values (Fst=0.0667; P=0.0037±0.0007).

Mitochondrial haplogroup composition

Mitochondrial haplogroup identified in the two population and the corresponding frequencies are shown in Table 2.

Results showed that both populations were entirely composed of Native American mitochondrial lineages. The four major Native American haplogroups (A, B, C, and D) were represented in both ethnic groups, but with a different frequency distribution. In the Kichwa sample, the most frequent haplogroups were A2* and its sublineage A2p with 49.23%; followed by D1f, identified in 40% of the population. Haplogroup C1* and its sublineages C1b and C1d were found in 7.69% of the individuals, whereas B4b was restricted to 3.08% of the Kichwa population.

In Mestizos, haplogroup A2* (and A2p) and haplogroup B (including B4b and B2h) were both detected in 33.33% of the population. Lower frequencies of haplogroups D1f and

Table 2 Mitochondrial DNA haplogroup percentages observed in Kichwa and Mestizo populations

Haplogroup	Kichwa	Mestizo
A2 ^a	21.54	26.19
A2p	27.69	7.14
B4b	3.08	30.95
B2h	_	2.38
C1 ^a	3.08	_
C1b	1.54	9.52
C1d	3.08	_
D1f	40.00	21.43
D4h3a	_	2.38

^a Further analysis of the coding region should be performed for a more accurate classification

C1b lineages were present, with 21.43% and 9.52%, respectively. Also the minor Native American haplogroup D4h3a was found in 2.38% of Mestizo individuals.

The lack of European and African contribution in maternal lineages can be explained through the historical background of these ethnic groups. In the Amerindian Kichwa population, no significant European or African contributions were expected to be found, as this group basically comprises individuals from different Amerindian tribes, which shared the same linguistic affiliation (Quechua). On the other hand, the Mestizo population mainly originated as a result of the admixture between European men and Native American women. Therefore, while male admixture is dominated by European Y chromosomes [8], the female gene pool shows the high influence of ancestral Native American mitochondrial lineages. This pattern is the result of the strong sexual asymmetry in the origin and ongoing history of the Mestizo population in Ecuador, which has also been described for other admixed populations from America [23, 24].

Furthermore, in this study, the comparison of the two Ecuadorian populations revealed genetic heterogeneity of the mtDNA gene pool, as demonstrated by the differences in the frequency of haplogroups and the presence of particular haplotypes. From the forensic perspective, the determination of population substructure in Ecuador supports the use of local population databases which reflect the genetic diversity existing among populations and/or ethnic groups of the country, whenever possible. The implementation of a single genetic database that wrongly assumes the homogeneity of the population could lead to erroneous interpretation of the weight of the evidence. The forensic field should not ignore population stratification, as assumptions about the homogeneity of pseudo-ethnic groups such as the "Hispanics" can have important repercussions on the statistical evaluation of the evidence in paternity testing and forensic casework.



Conclusions

The results of the present study corroborate the strong Amerindian influence in the Ecuadorian populations, which can be still observed in mitochondrial lineages despite the centuries of European presence and the extinction of a large number of Amerindian ethnic groups. Moreover, the results validate the usefulness of the analysis of the entire mtDNA control region in these populations, since it increases the discrimination power of mtDNA testing for forensic applications and adds valuable information to the phylogenetic interpretation of mtDNA haplotypes.

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Conflict of interest None.

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